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Degree Objective: Ph.D. Endocrinology and Reproductive Physiology

Background: BS Zoology, University of Wisconsin-Madison

MBA, California State University, San Marcos

Current Research Project:

Secretory proteins, which are predicted to account for nearly one third of all proteins that are translated, are delivered to the *cis*-Golgi apparatus from the endoplasmic reticulum (ER) via coat protein complex II (COPII)-coated vesicles. My lab has shown that a key regulator of the secretory pathway is Trk-fused gene (TFG), which localizes to sites of COPII vesicle biogenesis on the ER, facilitates the formation and anterograde transport of COPII vesicles, and maintains ER morphology. A major goal of my work is to better understand the mechanisms by which TFG functions in the early secretory pathway. Efficient protein secretion is required for normal endocrine signaling, as many hormones, including luteinizing hormone, follicle stimulating hormone, prolactin, and insulin, must be packaged into COPII-coated vesicles and transported through the Golgi apparatus into secretory granules, prior to their release into the bloodstream. Additionally, efficient protein secretion is critical during many stages of pregnancy. For example, decidual cells and the endometrial glands secrete growth factors and various other factors necessary for the implantation of the embryo. Thus, my work will have a broad impact on the fields of endocrinology and reproductive physiology.

Interestingly, mutations in TFG have been identified in several neurodegenerative disorders, including childhood-onset hereditary spastic paraplegia (HSP). HSP refers to a group of inherited disorders, characterized by progressive weakness and spasticity of the legs due to degeneration of upper motoneuron axons in the corticospinal tract. Individuals with HSP may also experience degradation of the optic nerve, ataxia, peripheral neuropathy, and deafness. My lab has shown that a mutation in TFG (p.R106C) causes a recessive form of HSP. To establish a tractable model for the study of this disease, I am using human iPSCs to determine the impact of the TFG p.R106C mutation on protein secretion and neuronal maintenance.

Human iPSCs can be derived from patient-specific somatic cells and can be directed to differentiate into all three germ layers of the human body. iPSCs are the ideal platform for extensively researching patient-specific diseases as well as *in vitro* testing of pharmaceuticals for these diseases. Although patient-derived iPSCs are often used to study the effects of genetic perturbations in culture, epigenetic effects can complicate or even confound the interpretation of results. Instead, I am taking advantage of a well-characterized iPSC line, and have used genome-editing approaches to create mutations in TFG that cause neurodegeneration. In this way, I was able to define the consequences of TFG mutations in a highly specific manner.



Grants Received:

NIH T32 Trainee; NIH Ruth L. Kirschstein National Research Service Award
NIH T32-HD041921 2015-2017.

Publications:

McMillan EL, Kamps AL, Lake SS, Svendsen CN, Bhattacharyya A, "Gene expression changes in the MAPK pathway in both Fragile C and Down syndrome human neural progenitor cells." American Journal of Stem Cells. June 2012.

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